

*Short Communication*

## Phytoplankton community structure at the eastern entrance of the Gulf of California during El Niño 2023

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**ABSTRACT.** This work aims to report the phytoplankton species composition and cell density in the Mazatlán coastal region (eastern entrance of the Gulf of California (GC), Mexico) during the strong El Niño event of 2023. Surface water samples (at 2 m depth) for phytoplankton cell determinations were collected in December of 2023, a month in which the numerical value of the Oceanic Niño Index was 2.0. The results showed a total of 197 species, including 113 diatoms (with a total cell density of 51,444 cells L<sup>-1</sup>), 76 dinoflagellates (with a total cell density of 31,260 cells L<sup>-1</sup>), four silicoflagellates (with a total cell density of 420 cells L<sup>-1</sup>), two cyanobacteria (with a total cell density of 2,020 cells L<sup>-1</sup>), one Euglenophyta (with a total cell density of 900 cells L<sup>-1</sup>) and one ciliate (with a total cell density of 7,820 cells L<sup>-1</sup>). The diatom *Dactyliosolen phuketensis* (B.G.Sundström) G.R. Hasle, 1996 and the dinoflagellate *Gyrodinium fusiforme* (Kofoid & Swezy, 1921) presented the highest cell densities with 4,100 and 5,900 cells L<sup>-1</sup>, respectively, species that have been previously reported in high abundances in Mexican waters in years of warming events, including El Niño. The results presented here contribute to understanding the effects of strong El Niño events on the phytoplankton community structure of the southeastern GC. This topic still needs to be fully addressed. This study also provides an update on the taxonomic lists available for the region of a group of organisms whose nomenclature has been constantly changing in recent years.

**Keywords:** phytoplankton species richness; phytoplankton abundance; El Niño; Mazatlán coastal region; Gulf of California

The last few years have seen extreme variability in the Earth's climate system. In fact, from the second half of 2020 to the first quarter of 2023, a strong La Niña event was recorded, which, in a quick transition, became an El Niño episode whose intensity can be compared to the strong event of 2015/2016. Data from the Oceanic Niño Index (ONI) showed that the last record of 2023 was 2.0, reflecting that a strong El Niño event was on record during December (NOAA 2024).

El Niño is one of the processes in the Earth's climate system that affect ecosystems and human populations

in several aspects, including agriculture, public health, energy generation, and consumption, tourism, the availability of freshwater, fishing, food security, and biodiversity, among others (McPhaden et al. 2021). El Niño has also been related to negative impacts on the phytoplankton community structure in the marine environment as a product of the advection of warm and oligotrophic water masses (Conde et al. 2018).

Phytoplankton is a pivotal group of organisms distributed in the euphotic layer of the oceans. Their photosynthetic processes prompt the release of oxygen

and the capture of CO<sub>2</sub> (Brierley 2017). Besides being the base of the food chain, phytoplankton is closely interconnected with the production in the higher trophic levels, both oceanic and coastal regions (Robicheau et al. 2022).

Coastal regions are sites recognized for their high diversity and as a place of refuge, growth, and feeding for numerous species (Hunter-Cevera et al. 2016). At the eastern entrance of the Gulf of California (GC), a sea recognized as one of the most biodiverse worldwide stands out in the Mazatlán coastal region (MCR).

As a part of the GC, the MCR is recognized for its high biological productivity levels, high rates of species richness (some of them endangered or threatened), high rates of endemisms, and for being the habitat of many migratory birds, mainly on the three islands that surround the region (Pájaros, Venados, and Chivos) (Hendrickx 1996, Esqueda-González et al. 2018). Hydrodynamically, the presence of tides, the alternation of a cyclonic/anticyclonic circulation pattern, the freshwater contribution of the Presidio River (with a discharge into the Pacific Ocean calculated at 65.8 million cubic meters annually), and an upwelling system make the MCR scientifically relevant. Besides, because of its geographical position, where the Pacific Ocean joins with the GC, the MCR is influenced by the presence of El Niño, a phenomenon that has received increasing attention in the last few years. Indeed, recent research unraveled some aspects related to the annual and interannual variability of sea surface temperature and phytoplankton biomass levels (expressed as chlorophyll-*a*), showing that El Niño has a recurrence interval of between three and four years inducing high sea surface temperature values (>31°C) and low chlorophyll-*a* concentrations (< 0.1 mg m<sup>-3</sup>) (Durán-Campos et al. 2023).

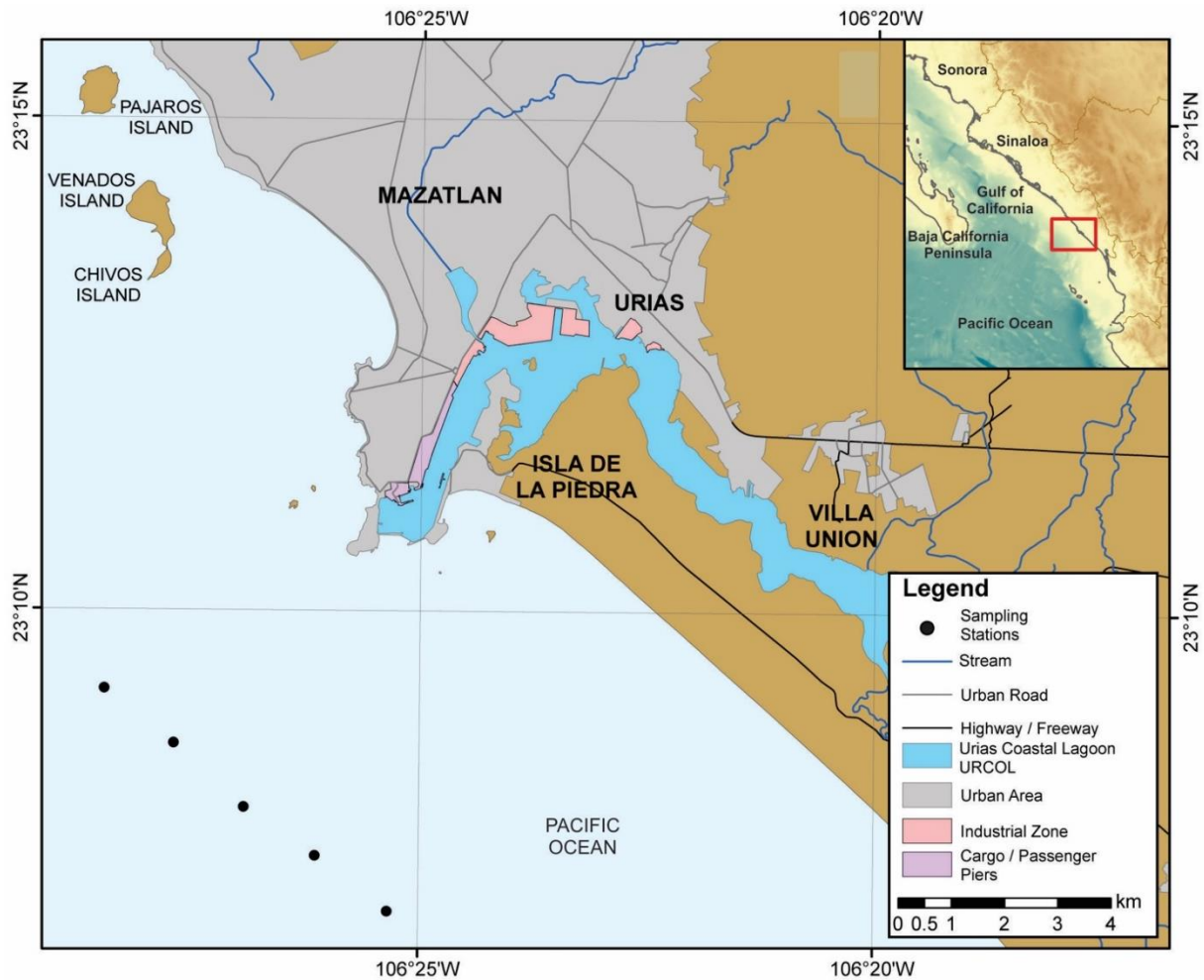
Studies on the phytoplankton community structure in the MCR have shown a high species richness and abundance. In one of the first taxonomic lists, more than 76 species were documented with marked seasonal variability, with the spring season having the highest number of species; in terms of dominance, diatoms have been reported as the species with the highest abundance relative to other species, constituting approximately 60% of the total abundance (Cortés-Altamirano & Pasten-Miranda 1982a,b, 1984, 1985).

In the MCR, species that generate harmful algal blooms have also been noticed. One of the earliest reports documented algal blooms generated by dinoflagellates (e.g. *Ceratium tripos* f. *ponticum* (Schiller, 1937) now called *Tripos muelleri* f. *ponticus*), *Prorocentrum triestinum* (J. Schiller, 1918), and

*Gymnodinium catenatum* (Graham, 1943)), by the diatom *Skeletonema costatum* (Cleve, 1873) and the ciliate (*Mesodinium rubrum* (Lohmann, 1908)); all of them associated with high mortality of fish, crustaceans, and annelids (Cortés-Altamirano 1987, Cortés-Altamirano et al. 2000, Hernández-Becerril et al. 2000). More recently, a trend toward an increase in toxic algal blooms frequency of the dinoflagellates *G. catenatum* and *Margalefidinium polykrikoides* (Margalef) F. Gómez, Richlen & D.M. Anderson, 2017 was documented with an increase in their duration (Cortés-Altamirano et al. 2019).

Although numerous studies have emerged in the scientific literature evaluating the phytoplankton community structure in the MCR, very few have directed their efforts on documenting the phytoplankton species composition and abundance under El Niño events. Hence, this short communication aims to report the phytoplankton community structure in December of 2023, a month characterized by a strong El Niño event. The above becomes more relevant when prediction models suggest that these events will be increasingly recurrent with larger impacts (Cai et al. 2014, Ying et al. 2022). Therefore, evaluating the impact of these events on phytoplankton community structure in regions recognized by their high biological productivity, such as the MCR, becomes scientifically necessary. For this, fieldwork was carried out on December 5, 2023, to collect water samples for cell determinations at five sampling points in the MCR (Fig. 1). Each collection site was reached with a handheld GPS (Garmin 64sx). Then, surface water (at 2 m depth) was collected with a UWITEC sampler bailer from which subsamples of 1 L were fixed immediately with a Lugol-acetate solution and kept in dark conditions until analyses.

The samples were analyzed immediately after collection using the Utermöhl method with 50 mL sedimentation columns (Edler & Elbrachter 2010). The sedimentation was in dark conditions for 24 h. Afterward, the phytoplankton organisms were identified to species level with a Carl Zeiss inverted microscope (Axiovert A1) using standard keys (e.g. Tomas 1997, Thronsen et al. 2003) and those keys published specifically for the Mexican Pacific Ocean and adjacent regions, such as the GC (e.g. Licea et al. 1995, Moreno et al. 1996, Esqueda-Lara & Hernández-Becerril 2010, Morquecho-Escamilla et al. 2016, Hernández-Becerril et al. 2021). Finally, the number of species was standardized to cell density units following standard protocols (Edler & Elbrachter 2010).



**Figure 1.** The study area, the Mazatlán coastal region, is at the eastern entrance of the Gulf of California, Mexico. The black dots represent the sampling stations.

The results showed a total of 197 species, including diatoms (113 species), dinoflagellates (76 species), silicoflagellates (4 species), Euglenophytes (1 species), ciliates (1 species), and cyanobacteria (2 species) (Table 1). In terms of cell density, the diatoms summarized 51,444 cells  $L^{-1}$ , the dinoflagellates 31,260 cells  $L^{-1}$ , the silicoflagellates 420 cells  $L^{-1}$ , cyanobacteria 2,020 cells  $L^{-1}$ , euglenophytes 900 cells  $L^{-1}$  and ciliates 7,820 cells  $L^{-1}$ .

Of the diatoms identified, *Dactyliosolen phuketensis* (B.G. Sundström) G.R. Hasle, 1996 was the most abundant species, reaching 4,100 cells  $L^{-1}$ , followed by *Leptocylindrus danicus* (Cleve, 1889) with 3,960 cells  $L^{-1}$ , while *Hemiaulus membranaceus* (Cleve, 1873), *Stigmaphora rostrata* (Wallich, 1860), *Asteromphalus heptactis* (Brébisson) Ralfs, 1861 among others, barely reached 20 cells  $L^{-1}$  (Table 1).

The dinoflagellate *Gyrodinium fusiforme* (Kofoid & Swezy, 1921) presented the highest abundance with 5,900 cells  $L^{-1}$ , while *Cucumeridinium lira* (Kofoid & Swezy) F. Gómez, P. López-García, H. Takayama & D. Moreira, 2015, *Grammatodinium tongyeonginum* (Z. Li & H.H. Shin, 2017), and *Proto-peridinium steinii* (Jørgensen) Balech, 1974 presented the lowest abundance (barely of 20 cells  $L^{-1}$ ). *Dictyocha californica* (Schrader & Murray 1985) was the most abundant silicoflagellate (300 cells  $L^{-1}$ ), and *Trichodesmium hildebrandtii* (Gomont, 1892) was the most abundant cyanobacteria. The Euglenophyta, *Euglena acusformis* (Schiller, 1926) reached 900 cells  $L^{-1}$ , and the ciliate *Mesodinium rubrum* (Lohmann, 1908) reached 7,820 cells  $L^{-1}$  (Table 1).

The fieldwork carried out in December 2023 allowed us to evaluate the phytoplankton community structure of the MCR during a strong El Niño event. To

**Table 1.** Phytoplankton species composition and their cell density (cells L<sup>-1</sup>) in the Mazatlán coastal region, eastern entrance of the Gulf of California, Mexico, during the strong El Niño event 2023.

Taxa	Cell density (cells L <sup>-1</sup> )
<b>Diatoms</b>	
<i>Actinocyclus octonarius</i> Ehrenberg, 1837	120
<i>Actinocyclus senarius</i> Ehrenberg, 1838	140
<i>Actinocyclus tenuissimus</i> Cleve, 1878	40
<i>Actinoptychus adriaticus</i> Grunow, 1863	820
<i>Alveus marinus</i> (Grunow) Kaczmarek & Fryxell, 1996	20
<i>Amphora costata</i> W.Smith, 1853	320
<i>Asterionellopsis glacialis</i> (Castracane) Round, 1990	160
<i>Asteromphalus arachne</i> (Brébisson) Ralfs, 1861	500
<i>Asteromphalus elegans</i> Greville, 1859	20
<i>Asteromphalus flabellatus</i> (Brébisson) Greville, 1859	100
<i>Asteromphalus heptactis</i> (Brébisson) Ralfs, 1861	20
<i>Asteromphalus ingens</i> Simonsen, 1974	20
<i>Asteromphalus roperianus</i> (Greville) Ralfs, 1861	20
<i>Azpeitia nodulifera</i> (A.W.F.Schmidt) G.A.Fryxell & P.A.Sims, 1986	160
<i>Bacteriastrum hyalinum</i> Lauder, 1864	100
<i>Bacteriastrum mediterraneum</i> Pavillard, 1916	1840
<i>Bellerophon malleus</i> (Brightwell) Van Heurck, 1885	40
<i>Cerataulina dentata</i> Hasle, 1980	180
<i>Cerataulina pelagica</i> (Cleve) Hendey, 1937	60
<i>Chaetoceros aequatorialis</i> Cleve, 1901	1580
<i>Chaetoceros affinis</i> Lauder, 1864	60
<i>Chaetoceros atlanticus</i> Cleve, 1873	40
<i>Chaetoceros aurivillii</i> Cleve, 1901	40
<i>Chaetoceros brevis</i> (Schütt, 1895)	180
<i>Chaetoceros coarctatus</i> Lauder, 1864	60
<i>Chaetoceros compressus</i> Lauder, 1864	640
<i>Chaetoceros costatus</i> Pavillard, 1911	80
<i>Chaetoceros curvisetus</i> Cleve, 1889	60
<i>Chaetoceros danicus</i> Cleve, 1889	160
<i>Chaetoceros decipiens</i> Cleve, 1873	320
<i>Chaetoceros densus</i> (Cleve) Cleve, 1899	160
<i>Chaetoceros denticulatus</i> H.S.Lauder, 1864	120
<i>Chaetoceros dictyota</i> Ehrenberg, 1844	20
<i>Chaetoceros didymus</i> Ehrenberg, 1845	40
<i>Chaetoceros diversus</i> Cleve, 1873	220
<i>Chaetoceros lacinosus</i> F.Schütt, 1895	20
<i>Chaetoceros mannaei</i> Li, Boonprakob, Moestrup & Lundholm, 2018	340
<i>Chaetoceros peruvianus</i> Brightwell, 1856	1500
<i>Chaetoceros protuberans</i> Lauder, 1864	40
<i>Chaetoceros rectus</i> Hernández-Becerril, 1992	300
<i>Chaetoceros rostratus</i> Ralfs, 1864	40
<i>Chaetoceros socialis</i> H.S.Lauder, 1864	580
<i>Chaetoceros teres</i> Cleve, 1896	1280
<i>Corethron hystrix</i> Hensen, 1887	140
<i>Corethron pennatum</i> (Grunow) Ostefeld, 1909	80
<i>Coscinodiscus asteromphalus</i> Ehrenberg, 1844	100
<i>Coscinodiscus centralis</i> Ehrenberg, 1839	100
<i>Coscinodiscus granii</i> L.F.Gough, 1905	160
<i>Coscinodiscus wailesii</i> Gran & Angst, 1931	220
<i>Cyclotella litoralis</i> Lange & Syvertsen, 1989	120

Continuation

Taxa	Cell density (cells L <sup>-1</sup> )
<b>Diatoms</b>	
<i>Cylindrotheca closterium</i> (Ehrenberg) Reimann & Lewin, 1964	360
<i>Dactyliosolen fragilissimus</i> (Bergon) Hasle, 1996	140
<i>Dactyliosolen mediterraneus</i> (H.Peragallo) H.Peragallo, 1892	1960
<i>Dactyliosolen phuketensis</i> (B.G.Sundström) G.R.Hasle, 1996	4100
<i>Detonula pumila</i> (Castracane) Gran, 1900	460
<i>Ditylum brightwellii</i> (T.West) Grunow, 1885	420
<i>Eucampia cornuta</i> (Cleve) Grunow, 1883	580
<i>Eupyxidicula palmeriana</i> (Greville) S.Blanco & C.E.Wetzel, 2016	20
<i>Eupyxidicula turris</i> (Greville) S.Blanco & C.E.Wetzel, 2016	180
<i>Guinardia flaccida</i> (Castracane) H.Peragallo, 1892	80
<i>Guinardia striata</i> (Stolterfoth) Hasle, 1996	480
<i>Gyrosigma spenceri</i> (Bailey ex Quekett) Griffith & Henfrey, 1856	20
<i>Haslea trompii</i> (Cleve) Simonsen, 1974	2240
<i>Haslea wawriake</i> (Husedt) Simonsen, 1974	300
<i>Helicotheca tamesis</i> (Shrubsole) M.Ricard, 1987	80
<i>Hemiaulus hauckii</i> Grunow ex Van Heurck, 1882	200
<i>Hemiaulus membranaceus</i> (Cleve, 1873)	20
<i>Hemiaulus sinensis</i> Greville, 1865	80
<i>Hemidiscus cuneiformis</i> Wallich, 1860	20
<i>Lauderia annulata</i> Cleve, 1873	20
<i>Leptocylindrus convexus</i> D.Nanjappa & A.Zingone, 2013	440
<i>Leptocylindrus danicus</i> Cleve, 1889	3960
<i>Lioloma pacificum</i> (Cupp) Hasle, 1996	100
<i>Lithodesmium undulatum</i> Ehrenberg, 1839	80
<i>Lithodesmium variabile</i> H.Tanako, 1979	40
<i>Meuniera membranacea</i> (Cleve) P.C.Silva, 1996	20
<i>Neodelphineis indica</i> (F.J.R.Taylor) Y.Tanimura, 1992	200
<i>Nitzschia bicapitata</i> Cleve, 1901	300
<i>Nitzschia ikeanae</i> Fryxell & H.Y.Lee, 1996	20
<i>Nitzschia longissima</i> (Brébisson ex Kützing) Grunow, 1862	2200
<i>Nitzschia sigma</i> (Kützing) W.Smith, 1853	20
<i>Odontella aurita</i> (Lyngbye) C.Agardh, 1832	40
<i>Planktoniella sol</i> (G.C.Wallich) Schütt, 1892	1420
<i>Pleurosigma acutum</i> (Norman ex Ralfs, 1861)	60
<i>Pleurosigma latum</i> Cleve, 1880	20
<i>Pleurosigma normanii</i> (Ralfs, 1861)	100
<i>Pleurosigma salinarum</i> (Grunow) Grunow, 1880	80
<i>Pseudo-nitzschia multistriata</i> (H.Takano) H.Takano, 1995	1500
<i>Pseudo-nitzschia pseudodelicatissima</i> (Hasle) Hasle, 1993	2100
<i>Pseudo-nitzschia pungens</i> (Grunow ex Cleve) Hasle, 1993	2340
<i>Pseudo-nitzschia roundii</i> Hernández-Becerril, 2006	3220
<i>Pseudo-nitzschia subfraudulenta</i> (Hasle) Hasle, 1993	140
<i>Pseudoguinardia recta</i> Stosch, 1986	200
<i>Ralfsiella smithii</i> (Ralfs) P.A.Sims, D.M.Williams & Ashworth, 2018	200
<i>Rhizosolenia bergonii</i> H.Peragallo, 1892	60
<i>Rhizosolenia pungens</i> Cleve-Euler, 1937	40
<i>Shionodiscus oestrupii</i> (Castracane) H.H.Gran, 1900	2920
<i>Skeletonema pseudocostatum</i> Medlin, 1991	1100
<i>Skeletonema tropicum</i> Cleve, 1900	740
<i>Stigmaphora rostrata</i> (Wallich, 1860)	20
<i>Tetramphora decussata</i> (Grunow) Stepanek & Kociolek, 2016	300
<i>Thalassionema bacillare</i> (Heiden) Kolbe, 1955	60

Continuation

Taxa	Cell density (cells L <sup>-1</sup> )
<b>Diatoms</b>	
<i>Thalassionema frauenfeldii</i> (Grunow) Tempère & Peragallo, 1910	680
<i>Thalassionema nitzschioides</i> (Grunow) Mereschkowsky, 1902	800
<i>Thalassiosira allenii</i> H. Takano, 1965	120
<i>Thalassiosira angustelineata</i> (A.W.F.Schmidt) G.Fryxell & Hasle, 1977	80
<i>Thalassiosira concaviuscula</i> Makarova, 1978	20
<i>Thalassiosira eccentrica</i> (Ehrenberg) Cleve, 1904	140
<i>Thalassiosira leptopus</i> (Grunow) Hasle & Fryxell, 1977	20
<i>Thalassiosira punctifera</i> (Grunow) Fryxell, Simonsen & Hasle, 1974	20
<i>Thalassiosira tenera</i> Proshkina-Lavrenko, 1961	180
<i>Triceratium formosum</i> Brightwell, 1856	80
<i>Trieres mobiliensis</i> (Bailey) Ashworth & E.C.Theriot, 2013	20
<b>Dinoflagellates</b>	
<i>Achradina pulchra</i> Lohmann, 1903	200
<i>Akashiwo sanguinea</i> (K.Hirasaka) Gert Hansen & Moestrup, 2000	860
<i>Alexandrium minutum</i> Halim, 1960	200
<i>Alexandrium monilatum</i> (J.F.Howell) Balech, 1995	840
<i>Alexandrium tamarense</i> (Lebour) Balech, 1995	80
<i>Azadinium caudatum</i> (Halldal) Nézan & Chomérat, 2012	280
<i>Azadinium poporum</i> Tillmann & Elbrächter, 2011	800
<i>Azadinium spinosum</i> Elbrächter & Tillmann, 2009	1100
<i>Blepharocysta denticulata</i> D.-S.Nie, 1939	1620
<i>Blepharocysta paulsenii</i> Schiller, 1937	40
<i>Boreadinium breve</i> (T.H.Abé) Sournia, 1984	20
<i>Centrodinium pulchrum</i> Bohm, 1933	20
<i>Ceratocorys armata</i> (Schütt) Kofoid, 1910	20
<i>Corythodinium biconicum</i> (Kofoid) F.J.R.Taylor, 1976	20
<i>Corythodinium mucronatum</i> (B.Hope) F.Gómez, 2017	20
<i>Cucumeridinium coeruleum</i> (Dogiel) F.Gomez, P.López-García, H.Takayama & D.Moreira, 2015	80
<i>Cucumeridinium lira</i> (Kofoid & Swezy) F.Gómez, P.López-García, H.Takayama & D.Moreira, 2015	20
<i>Diplopelta asymmetrica</i> (Mangin) M.Lebour ex Balech, 1988	1920
<i>Diplopsalopsis orbicularis</i> (Paulsen) Meunier, 1910	80
<i>Fragilidium mexicanum</i> Balech, 1988	20
<i>Gonyaulax fusiformis</i> H.W.Graham, 1942	20
<i>Grammatodinium tongyeonginum</i> Z.Li & H.H.Shin, 2017	20
<i>Gymnodinium catenatum</i> H.W.Graham, 1943	60
<i>Gymnodinium impudicum</i> (S.Fraga & I.Bravo) Gert Hansen & Moestrup, 2000	600
<i>Gyrodinium fusiforme</i> Kofoid & Swezy, 1921	5900
<i>Heterocapsa orientalis</i> Iwataki, Botes & Fukuyo, 2003	80
<i>Kapelodinium vestifici</i> (Schütt) Boutrup, Moestrup & Daugbjerg, 2016	40
<i>Karenia bicuneiformis</i> Botes, Sym & Pitcher, 2003	560
<i>Karenia brevisulcata</i> (F.H. Chang) Gert Hansen & Moestrup, 2000	1100
<i>Lingulodinium polyedrum</i> (F.Stein) J.D. Dodge, 1989	1980
<i>Nematodinium armatum</i> (Dogiel) Kofoid & Swezy, 1921	20
<i>Noctiluca scintillans</i> (Macartney) Kofoid & Swezy, 1921	20
<i>Ornithocercus heteroporus</i> Kofoid, 1907	40
<i>Oxytoxum sceptrum</i> (F.Stein) Schröder, 1900	700
<i>Oxytoxum variabile</i> J.Schiller, 1937	160
<i>Phalacroma argus</i> F.Stein, 1883	20
<i>Prorocentrum gracile</i> F.Schütt, 1895	800
<i>Prorocentrum koreanum</i> M.-S.Han, S.Y.Cho & P.Wang, 2016	560
<i>Prorocentrum lenticulatum</i> (Matzenauer) F.J.R.Taylor, 1976	120
<i>Prorocentrum mexicanum</i> Osorio-Tafall, 1942	140

Continuation

Taxa	Cell density (cells L <sup>-1</sup> )
<b>Dinoflagellates</b>	
<i>Prorocentrum obtusidens</i> J.Schiller, 1928	400
<i>Prorocentrum robustum</i> Osorio-Tafall, 1942	60
<i>Prorocentrum sigmoides</i> Böhm, 1933	40
<i>Protoceratium reticulatum</i> (Claparède & Lachmann) Bütschli, 1885	40
<i>Protoperidinium acutum</i> (G.Karsten) Balech, 1974	40
<i>Protoperidinium bispinum</i> (J.Schiller) Balech, 1974	80
<i>Protoperidinium brochii</i> (Kofoid & Swezy) Balech, 1974	20
<i>Protoperidinium claudicans</i> (Paulsen) Balech, 1974	40
<i>Protoperidinium conicum</i> (Gran) Balech, 1974	40
<i>Protoperidinium consimile</i> (T.H.Abé) Balech, 1994	60
<i>Protoperidinium corniculum</i> (Kofoid & J.R.Michener) F.J.R.Taylor & Balech, 1988	160
<i>Protoperidinium depressum</i> (Bailey) Balech, 1974	20
<i>Protoperidinium divergens</i> (Ehrenberg) Balech, 1974	20
<i>Protoperidinium obtusum</i> (Karsten) Parke & J.D.Dodge, 1976	2380
<i>Protoperidinium pentagonum</i> (Gran) Balech, 1974	40
<i>Protoperidinium perplexum</i> (Balech) Balech, 1974	20
<i>Protoperidinium punctulatum</i> (Paulsen) Balech, 1974	2200
<i>Protoperidinium pyrum</i> (Balech) Balech, 1974	40
<i>Protoperidinium robustum</i> (Meunier) Hernández-Becerril, 1991	1260
<i>Protoperidinium saltans</i> (Meunier) Balech, 1973	20
<i>Protoperidinium simulum</i> (Paulsen) Balech, 1974	40
<i>Protoperidinium steinii</i> (Jørgensen) Balech, 1974	20
<i>Protoperidinium subinerme</i> (Paulsen) A.R.Loeblich III, 1969	40
<i>Protoperidinium tuba</i> (J.Schiller) Balech, 1974	1160
<i>Ptychodiscus noctiluca</i> Stein, 1883	20
<i>Pyrocystis pseudonociluca</i> Wyville-Thompson, 1876	200
<i>Scrippsiella acuminata</i> (Ehrenberg) Kretschmann, Elbrächter, Zinssmeister, S. Soehner, Kirsch, Kusber & Gottschling, 2015	20
<i>Scrippsiella spinifera</i> G.Honsell & M.Cabrini, 1991	280
<i>Torquentidium convolutum</i> (Kofoid & Swezy) H.H.Shin, Z.Li, K.W.Lee & K.Matsuoka, 2019	40
<i>Torquentidium helix</i> (Lemmermann) H.H.Shin, Z.Li, K.W.Lee & K.Matsuoka, 2019	160
<i>Triadinium polyedricum</i> (Pouchet) J.D.Dodge, 1981	20
<i>Tripes balechii</i> (Meave, Okolodkov & M.E.Zamudio) F.Gómez, 2013	80
<i>Tripes falcatus</i> (Kofoid) F.Gómez, 2013	120
<i>Tripes furca</i> (Ehrenberg) F.Gómez, 2013	840
<i>Tripes gravidus</i> (Gourret) F.Gómez, 2013	40
<i>Tripes trichoceros</i> (Ehrenberg) F.Gómez, 2013	20
<b>Silicoflagellates</b>	
<i>Dictyocha calida</i> Poelchau, 1976	20
<i>Dictyocha californica</i> Schrader & Murray, 1985	300
<i>Dictyocha fibula</i> Ehrenberg, 1839	60
<i>Octactis octonaria</i> (Ehrenberg) Hovasse, 1946	40
<b>Euglenophyta</b>	
<i>Euglena acusformis</i> J.Schiller, 1926	900
<b>Ciliates</b>	
<i>Mesodinium rubrum</i> Lohmann, 1908	7820
<b>Cyanobacteria</b>	
<i>Trichodesmium hildebrandtii</i> Gomont, 1892	1360
<i>Trichodesmium thiebautii</i> Gomont, 1890	660

the best of our knowledge, in the MCR, there are very few reports on the phytoplankton species richness and their abundance during El Niño events. Therefore, it becomes complicated to establish robust comparisons. However, some studies focused mainly on evaluating the generation of harmful algal blooms in the region suggest that El Niño may have a direct effect on the presence and intensity of the blooms due to the advection of warm water masses that, in turn, weakens the upwelling processes that occur there (Cortes-Altamirano 1987).

In our study, diatoms presented the highest number of species, agreeing with previous reports that pointed to this group as dominant in the phytoplankton community structure in the MCR. Indeed, some studies have reported that diatoms usually dominate around 60% of the total (e.g. Cortes-Altamirano & Pasten-Miranda 1985), as was our case.

In our study, the diatom *Dactyliosolen phuketensis* presented the highest abundance. This species has been reported in Mexican waters in years with warming events, such as the case of El Blob 2013/2015 (a sudden overheating of surface waters in the Pacific Ocean that occurred in that period whose causes are not yet fully understood) and El Niño 2015/2016 (Jiménez-Quiroz et al. 2019) and in the Acapulco Bay (Mexican Pacific) in March of 2010. This month presented El Niño conditions (Meave-del-Castillo et al. 2012). Concomitantly, studies carried out in the southern GC (González-López & Siqueiros-Beltrones 1990), in the western coast of South America (Conde et al. 2018), and in the southwestern Atlantic Ocean (Barrera-Alba et al. 2019) have documented that species of the genus *Dactyliosolen* tend to be indicators of warming events associated with the presence of El Niño events; even, blooms of species of this genus have been related to El Niño conditions (González-Narváez et al. 2021), which could explain the high cell density observed in our study.

*Gyrodinium fusiforme* was the most abundant dinoflagellate in our study (with 5,900 cells L<sup>-1</sup>). This species has been reported in high cell densities in Mexican waters during El Niño events, particularly in Acapulco Bay (Mexican Pacific) (Meave-del-Castillo et al. 2012) and in Concepción Bay (GC) (Verdugo-Díaz et al. 2010). Species of the genus *Gyrodinium* have also been reported with high cell densities related to El Niño events in different environments around the world. Indeed, in northern Chile, increases in species of this genus were reported during El Niño 1982/1983 (Avaria & Muñoz 1987). In Hong Kong, an atypical bloom of *Gyrodinium aureolum* was reported during El

Niño 1997/1998 (Yin et al. 1999), while high cell densities of *Gyrodinium acutum* have been reported in Ecuador (Eastern Pacific Coast) during El Niño events (Torres et al. 2019).

Another species observed with relatively high cell density was the ciliate *Mesodinium rubrum*, previously reported in the MCR as bloom-forming algae (Cortes-Altamirano 1987). Numerous reports in the scientific literature on the presence of this species in El Niño events, including the Peruvian coast (Sánchez et al. 2021), the Ecuadorian coast (Conde et al. 2018), and the Galápagos Islands (Carnicer et al. 2019), agree with our observations.

In summary, the results presented here contribute to understanding the effects of strong El Niño events on the phytoplankton community structure in the MCR. This study also provides an update of the taxonomic lists available for the region of a group of organisms whose nomenclature has been constantly changing in recent years. Besides, the taxonomic list presented here represents a reference to the phytoplankton community structure under strong El Niño events; the last could act as a baseline for future studies in the region under different events, whether El Niño or La Niña.

#### Credit author contribution

E. Durán-Campos: conceptualization, validation, sampling, species identification, formal analysis, writing-original draft; C.M. Torres-Martínez: validation, sampling, species identification, formal analysis, writing-original draft; E. Coria-Monter: conceptualization, validation, sampling, species identification, formal analysis, writing-original draft; D.A. Salas de León: conceptualization, validation, formal analysis, funding acquisition, writing-original draft; M.A. Monreal-Gómez: conceptualization, validation, formal analysis, funding acquisition, writing-original draft. All authors have read and accepted the published version of the manuscript.

#### Conflict of interest

The authors declare no potential conflict of interest in this manuscript.

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